How Did It Get Here? First Record of *Trochamminita irregularis*, a Cosmopolitan Estuarine Organic-Cemented Agglutinated Foraminifer, in South-West Western Australia

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Abstract

The cosmopolitan organic-cemented agglutinated foraminifer *Trochamminita irregularis* previously identified in Australian salt marshes of the Gippsland Lakes in Victoria and Little Swanport Estuary in Tasmania, has been recognised for the first time in Western Australia in the Hay River connected to Wilson Inlet, in the south-west of the State. The irregular test and chamber shapes may be related to the species mode of living restricted to the upper-marsh or river-margin environments in organic-rich sediment associated with filamentous rhizomes/stolons of marsh vegetation. Morphological variation may also be due to highly variable seasonal to daily environmental changes ranging from tidal variation, salinity changes (within a hyposaline range) and variable river flow. *Trochamminita*'s fragmented global distribution could likely be attributed to attachment to migratory waterbirds dispersing it along major flyways such as the East Australasian Flyway where Australia acts as one of the main foraging and breeding ground for these birds. The discovery of the species in the Hay River allows some preliminary investigations of the micro-living habitat of *Trochamminita* and its potential means of dispersion. Molecular and more in-depth ecological studies on living *T. irregularis* are required to more fully understand this global and ecologically significant marsh indicator species.

INTRODUCTION

Salt marshes and surrounding coastal estuarine vegetation act as dynamic intertidal ecosystems located at the interface between terrestrial and marine environments (Brearley, 2005; Reed et al., 2009). Present along most estuaries worldwide, these muddy, anaerobic and organicrich microhabitats are characterized by halophytic vegetation – withstanding fluctuations between brackish and freshwater conditions caused by daily and seasonal change. These habitats not only are recognised for their extensive ecosystem services, serving as natural buffers against coastal erosion and flooding, but are also home to a diversity of micro- to macro-organisms, encompassing amphipods, foraminifers, bivalves and gastropods (Murray, 2014; Pennings & Bertness, 2001; Ross et al., 2009). The complex zone of rhizomes and stolons at the edge of the marsh comprises an intricate matrix, that accounts for a highly interconnected web of biological and physical processes promoting nutrient cycling and oxygen regulation (Vernberg, 1993).

Trochamminita irregularis (Cushman and Brönnimann) is one of two species known within its genus, belonging among the organic-cemented agglutinated foraminifera that do not secrete a calcium-carbonate component in their test. These differ from carbonate-cemented agglutinated foraminifera and other groups that produce calcium carbonate through biomineralization. The species is characterised by highly variable whorl arrangements, often reaching up to four in adult specimens. Its chambers initially coil in a planispire, with the test axis shifting and twisting during later development. Trochamminita irregularis differs from the other species within its genus, Trochamminita salsa (Cushman and Brönnimann), by its asymmetrical chamber arrangement, and its wider adaptability to salinity change, compared to a more saline condition for T. salsa. Trochamminita irregularis has a broad and cosmopolitan distribution, spanning diverse geographic and ecological regions (Fig. 1, Table 1). In North America, it is found along the Atlantic and Pacific coasts, including upper

marshes of Oregon (e.g., Salmon River and South Slough), Texas (Trinity River), North Carolina (Outer Banks), and Florida (Everglades). It also occurs in estuarine systems like the Pascagoula River in Mississippi and Bayou Lafourche in Louisiana. Further south, in South America, it inhabits mangrove and marsh systems in Brazil (Caeté River) and French Guiana (Kaw estuary). Its presence in Europe is notable in inland saline springs of Germany and upper marshes in Iceland and Portugal (e.g., Caminha Marsh). In the Southern Hemisphere, *T. irregularis* is distributed across various wetlands and marshes in Australia (e.g., Little Swanport in Tasmania) and New Zealand (e.g., Mokomoko Inlet and Pauatahanui Inlet). It has also been recorded in the saline environments of Lake Verlorenvlei, South Africa, in the estuarine lagoon of Seitu in Malaysia and in the tropical mangroves of Saint Vincent Bay, New Caledonia.

This preliminary study aims to (i) document the first discovery of *Trochamminita irregularis* in a southwestern Australian estuary, (ii) describe its micro-living habitat in Hay River, Wilson Inlet; and (iii) consider the pathways of its global distribution.

NEW LOCALITY OF TROCHAMMINITA IRREGULARIS

Wilson Inlet is a seasonally open, broad tidal inlet located on the southern coast of Western Australia (Ranasinghe & Pattiaratchi, 1999; Brearly 2005). It is a shallow estuarine basin with an area of 48 km² and a mean depth of 1.8 m, exhibiting freshwater conditions in the winter months and brackish conditions in the summer (Lukatelich et al., 1987; Twomey & Thompson, 2001). The Hay River (Fig. 2a) is one of three main tributaries (with salinity varying from freshwater 8-10 ppt in winter to brackish 27-28 ppt in the summer months) flowing into the inlet, and it accounts for 65% of the annual streamflow into the estuary (Ranasinghe & Pattiaratchi, 1999). The Hay River catchment features two distinct landform types: with (1) a western re-



Figure 1. Distribution of *Trochamminita irregularis*. a) Global distribution (white circles, number referenced in <u>Table 1</u>) in relation to the eight broad major avian flyways including Pacific Americas, Central America, Atlantic Americas, East Atlantic, Central Asia, Black Sea-Mediterranean, East Asia-East Africa, and East Australasia (adapted from Bamford et al., 2008); b) enlarged view with average annual isohyets (Bureau of Meteorology, 2024) of the southern Australian coastline showing Locality 23 (starred, studied area) near mouth of Hay River flowing into Wilson Inlet, and Gippsland Lakes and Little Swanport Estuary, the only other localities where *T. irregularis* has been recorded in Australia.

gion characterized by a rolling, dissected lateritic terrain that transitions into lateritic uplands (Lukatelich et al., 1984). In contrast, (2) the eastern portion of the catchment, which also includes the Sleeman River catchment, consists of lateritic sandplain swamps and plains interspersed with gravelly ridges (Lukatelich et al., 1984). The low relief in this area results in swampy flats surrounding the stream channels (<u>Fig. 2b</u>). The soils are primarily leached sands and yellow-mottled soils, which support jarrah scrub, and sandplain heaths (Lukatelich et al., 1984).

Trochamminita irregularis is found in low abundance in shoreline environments near the confluence of Hay River and Wilson Inlet at coordinates 34.970089°S, 117.4692702°E (Fig. 2c). It forms <20% of the marsh foraminiferal fauna in a systematic pick of dead and live individuals, alongside *Ammonia haigi* Haywood and Holzmann, *Trochammina inflata* (Montagu), and *Elphidium* sp., and lives among riparian vegetation including *Sarcocornia quinqueflora* Bunge ex Ung.-Sternb. (Fig. 2d), *Baumea* sp. and *Juncus* sp. (Fig. 2e). The substrate is composed of fine silty mud with very rare shelly material. Morphological variability (Fig. 3a–1) of the test of *T. irregularis* could likely be shaped by the intertwining pattern of above-ground stolons at the base of the vegetation. The multiple apertures and their varying position either at the base of the ultimate chamber or areal in the apertural face of this chamber (Figs. 3j, k, 1) may be a potential morphological adaptation to maximise food uptake and to facilitate the protrusion of the pseudopodia in 3-dimensions, while attached to the filamentous stolons/rhizomes. In the slightly deeper part of the Hay River (>0.5 m water depth), the seagrass *Ruppia megacarpa* Manson is found in occasional patches growing in a mix of sandy and silty mud. *Trochamminita irregularis* was not found in this habitat.

Table 1. Selected worldwide sites with Trochamminita irregularis, including habitat zones and records of associated foraminifers.

No	Localities	Countries	Habitat zone	Other foraminifers	References
1	Salt Lake, Honolulu Hawaii	USA	Upper marsh	Ammonia tepida, Haplophragmoides manilaensis, Milliamina fusca	Resig, 1974
2	Port Alberni, British Columbia	USA	Upper marsh	Entzia macrescens, Trochammina inflata, Haplophragmoides wilberti, M. fusca	Clague et al., 1994; Ozarko et al., 1997
3	Salmon River, Oregon	USA	Upper marsh	Balticammina pseudomacrescens, Haplophragmoides wilberti, Trochammina inflata, Entzia macrescens and M. fusca	Hawkes et al., 2010; Milker et al., 2015
4	South Slough, Oregon	USA	Upper marsh	B. pseudomacrescens, H. wilberti, T. inflata, E. macrescens, M. fusca	Hawkes et al., 2010; Milker et al., 2015
5	Tijuana River, California	USA	Upper marsh	E. macrescens, T. inflata, B. pseudomacrescens, H. wilberti	Avnaim-Katav et al., 2017, 2023
6	Trinity River, Texas	USA	Upper marsh	E. macrescens, M. fusca, T. inflata, H. wilberti.	Brann, 1969
7	Pascagoula River and Grand Bay, Mississippi Sound	USA	Upper marsh	A. tepida, Cribroelphidium poeyanum, C. excavatum, and Paratrochammina simplissima, Ammotium salsum, Ammobaculites exiguus, M. fusca, Arenoparrella mexicana, E. macrescens, Pseudothurammina limnetis.	Haller et al., 2019
8	Bayou Lafourche, Louisiana	USA	Upper marsh	T. inflata, Ammotium crassus, Ammonia parkinsoniana, Polysaccammina ipobalina, M. fusca	Dreher, 2006
9	Florida Everglade	USA	Upper marsh	E. macrescens, M. fusca, H. wilberti, A. mexicana, Ammonia spp., Elphidium spp., A. tepida, Helenina anderseni, T. inflata	Z. R. Verlaak & Collins, 2021; Z. R. F. Verlaak, 2019
10	Pamlico Sound and Currituck Sound coasts of North Carolina's Outer Banks	USA	Upper marsh	A. mexicana, H. wilberti, E. macrescens, T. inflata, M. fusca, Tiphotrocha comprimata	Culver & Horton, 2005; Robinson & McBride, 2006
11	Maracas Bay River, north coast	Trinidad	Shallow tributary drain of the Maracas Bay River.	H. manilaensis, H. wilberti, Trochamminita salsa, Trochammina laevigata, T. comprimata, Siphotrochammina lobata, A. mexicana	Cushman & Brönnimann, 1948; L. Laut et al., 2017; Saunders, 1957
12	Kaw estuary and Lagoon of Montjoly	French Guinea	Mangrove swamps	A. tepida, A. parkinsoniana, Cribroelphidium spp., M. fusca	Debenay et al., 2002
13	Caete River	Brazil	Upper marsh and mangroves	T. paranaguaensis, D. urceolata, D. oblonga	L. L. M. Laut et al., 2016
14	Rio Itapanhaú	Brazil	Upper marsh	Not described	Moreno, 2004
15	Valdivia- Tornagaleones estuary	Chile	Upper marsh	M. fusca, T. salsa, Haplophragmoides spp., Trochammina squamata	Jennings et al., 1995
16	Melabakkarin vicinity of Reyjavik	Iceland	Upper marsh	Haynesina orbicularis, E. macrescens	Lübbers & Schönfeld, 2018
17	Bay of Tümlau, Schleswig-Holstein	Denmark	Upper marsh	<i>B.pseudomacrescens, T. inflata, E. macrescens</i> , and monothalamous <i>Ovammina opaca</i>	Bunzel et al., 2013
18	Saxony-Anhalt and Thuringia	Central Germany	Saline springs	T. salsa, E. macrescens, M. fusca, S. lobata, H. manilaensis, H. wilberti	Lehmann, 2000; Milker et al., 2023
19	Caminha marsh, Minho River	Portugal	Upper marsh	H. manilaensis	Fatela et al., 2014; Semensatto, 2020
20	Aveiro	Portugal	Upper marsh	Rotaliammina concava, Lepidodeuterammina ochracea, Quinqueloculina seminula, Gavelinopsis praegeri, Paratrochammina haynesi, Remaneica helgolandica, Remaneicella gonzalezi	Martins et al., 2019
21	Lake Verlorenvlei	South Africa	Enclosed lake, upper marsh	Haplophragmoides sp., Trochammina inflata, Milliamina earlandi, Ammonia parkinsoniana, Ammonia tepida	Fürstenberg et al., 2017
22	Setiu estuary–lagoon system, Terengganu	Malaysia	Upper marsh/ wetland	Ammobaculites exiguus, Bruneica clypea, Caronia exilis, Haplophragmoides wilberti, Siphotrochammina lobata, and Trochammina inflata	Culver et al., 2015
23	Wilson Inlet	Australia	Upper marsh, restricted Hay River	Ammonia haigi, T. inflata, Elphidium spp.	This study
24	Gippsland Lakes	Australia	At margin	M. fusca, A. salsum, Reophax barwonensis, "Martinotiella cf.	Apthorpe, 1980

	(Lake Wellington)		oflake	communis" (= ?Scherochorella), T. inflata	
25	Little Swanport, Tasmania	Australia	Upper marsh	T. inflata, E. macrescens, T. salsa, M. fusca, Elphidium sp., H. wilberti, Quinqueloculina sp.	Callard et al., 2011; Gehrels et al., 2012; Williams et al., 2021
26	Port Pegasus "Pikihatiti"	New Zealand	Upper marsh	T. salsa, T. inflata, H. wilberti, M. fusca, Ammonia spp., E. macrescens	Hayward et al., 1994
27	Mokomoko Inlet	New Zealand	Upper marsh	Trochamminita spp., H. wilberti, M. fusca	Garrett et al., 2022
28	Pauatahanui Inlet, southern North Island	New Zealand	Upper marsh	T. salsa, Pseudotrochamminita malcomi, Polysaccammina ipohalina	King, 2021
29	Saint Vincent Bay	New Caledonia	Mangroves	M. fusca, A. mexicana, H. wilberti, T. inflata, Trochammina patensis	Debenay et al., 2015

DISCUSSION

Associated foraminifers with *Trochamminita irregularis*

Worldwide species associations

Trochamminita irregularis commonly coexists with a variety of foraminiferal species that inhabit similar marsh, estuarine, and mangrove ecosystems. In temperate and subtropical marshes, it is often found alongside species such as Entzia macrescens (Brady), often referred to Jadammina, and Miliammina fusca (Brady), which are wellknown indicators of high-marsh environments (Table 1). The association with Trochammina inflata, another marsh specialist, is consistent in regions like Oregon, New Zealand, and South America. In saline springs, it is found with E. macrescens and Siphotrochammina lobata Saunders, highlighting its adaptability to high salinity and fluctuating environmental conditions. In tropical and subtropical ecosystems, T. irregularis cohabits with Arenoparrella mexicana (Kornfeld), Haplophragmoides wilberti Anderson, and species of Ammonia (Table 1). Other notable cohabitants include Balticammina pseudomacrescens Brönnimann, Lutze & Whittaker in northern marshes (e.g., Oregon) and Trochamminita salsa, a species found in South America, Germany, and New Zealand. In Portugal, Australia, and Malaysia, T. irregularis is also associated with species often referred to as Quinqueloculina "seminula" (Linnaeus), Paratrochammina haynesi (Atkinson), and Polysaccammina ipohalina Scott, which are indicative of brackish to marine transitions. By understanding these co-occurrences, it does underline the ecological versatility and habitat diversity in which T. irregularis evolved but also raises significant questions about dispersal of the species.

Hay River species association

Based on previous and continuing studies on Western Australian estuaries, the Hay River site described here is the only locality where *Trochamminita irregularis* has been found in the State (Table 2). The dominant foraminiferal species at the Hay River study site are the calcareous trochospiral *Ammonia haigi* and the organic-cemented agglutinated trochospiral *Trochammina inflata*. Apart from the Hay River site, *T. inflata* is known from 12 of the other 28 world-wide *T. irregularis* sites listed on Table 1, and species of *Ammonia* were recorded from six of the sites. These species are known from many of the other estuaries in south-west Western Australia. Questions exist, that require much further work and cannot be answered in this short note, about why different species are associated with *Trochamminita* in the marsh associations at different localities (Table 1), and why *T. irregularis* has not been found at other sites in Western Australia containing associated marsh foraminifers (based on extensive unpublished work on most estuaries from Esperance in the east to Carnarvon in the northwest).

Biogeographical fragmentation: potential means of distribution

The global distribution of *T. irregularis* is highlighted by a fragmented pattern ranging from tropical to temperature regions, and by differences in associated species almost all of which are confined to marshes and river-margin situations under mainly brackish water conditions. Most estuaries are disconnected spatially and in geological time. This is very apparent between the two known occurrences of *Trochamminita irregularis* on the southern coastline of Australia, Gippsland Lakes in the east and Hay River in the west, with about 1500 km of the >2500 km shoreline between these sites lacking river inflow and estuaries.

Dispersal of estuarine foraminifera has been suggested to occur by different means including anthropogenic interventions (ballast water, e.g., Tremblin et al., 2021), or from natural mechanisms (drifting of foraminiferal propagules in the current, e.g. Alve, 1999, and rafting e.g., Finger, 2018). Bird-mediated dispersal of riverine invertebrates including foraminifera has been suggested in the literature through ectozoochory and endozoochory, where the biota get into feathers, feet or directly in the birds digestive systems (Coughlan et al., 2017; Riedel et al., 2011). Dispersal events by birds may be rare but because of the large migratory waterbird populations and the countless days of migration from their departure point to their destination, the cumulative rate of rare dispersals make these significant.

In the Australian context, dispersal of estuary-bound foraminifers must have been across large terrestrial spaces (either north-south or east-west) or over large tracts of ocean surrounding the continent. Australia is located at the edge of one of eight major flyways for migratory land and waterbirds (Fig. 1). This flyway accounts for about 700 species of birds flying from as far away as Siberia in the north and to New Zealand in the South. In their foraging, waterbirds may unintentionally transport T. irregularis through their feathers, feet, or digestive systems as they migrate across vast distances. Five species of waterbirds including the Eurasian Coot (Fulica atra), Red-necked Stint (Calidris ruficollis), Silver Gull (Chroicocephalus novaehollandiae), Fairy Tern (Sternula nereis) and Caspian Tern (Hydroprogne caspia), share the three localities where Trochamminita irregularis has been recorded in Australia (Birdlife Western Australia, 2024; Bryant, 2002; Hansen et al., 2024). Under extreme conditions foraminiferal protoplasm can remain viable for long periods of time, up to two years, as described by Alve and Goldstein (2010). It has also been shown from fish-gut contents that more than 20% of foraminiferal protoplasm is still viable even after days of being digested (Guy-Haim et al., 2017). In the case of *T. irregularis*, their asexual reproduction, as well as their flexible, imperforate, and highly variable tests built on an organic template, and their ability to construct their tests using surrounding fine siliceous sand grains, probably make them likely to quickly regener-



Figure 2. Drone images of the Hay River connected to Wilson Inlet. a) view of Hay River mouth flowing into Wilson Inlet (yellow dots – localities of sampled *Trochamminita irregularis*); b) view of Hay River upstream from sampled sites; c) close-up view of river channel adjacent to the sampled sites; d, e) common salt-marsh vegetation including (d) *Sarcocornia quinqueflora* Bunge ex Ung.-Sternb. and (e) *Baumea* sp. and *Juncus* sp., where *T. irregularis* lives.

ate a population when an environmentally suitable habitat is encountered. The other taxa associated with *T. irregularis* could have been transported via the same mechanisms. The occurrence of *T. irregularis* in Hay River adds to the presently known record of the species distribution and emphasizes the ecological importance of studying saltmarsh environments in Western Australia and elsewhere and using foraminifers as markers for not only possible recent introductions, but also as environmental proxies.

Future work should involve (1) genetic analysis to understand the connectivity of the various estuarine *Trochamminita* populations worldwide including in areas of overlap of the migratory flyways, as well as the genetic drift along possible migratory pathways; (2) seasonal ecological monitoring of *T. irregularis* in the Hay River, to note

species dynamics under varying climatic conditions; and (3) analyses of mud detritus attached to feathers and feet as well as the faecal content of migratory birds to determine if attached mud and faeces could facilitate the foraminiferal migration.

CONCLUSIONS

Trochamminita irregularis, a species recognized for its global distribution, previously recorded in Australia only in the Gippsland Lakes (Victoria) and Little Swanport Estuary in Tasmania, has now been identified in the Hay River of Wilson Inlet, located in south-western Australia. This species is distinguished by its variable chamber mor-



Figure 3. *Trochamminita irregularis* (Cushman and Brönnimann) from the Hay River sites shown in Fig. 2, emphasizing the morphological variation in test shape, agglutination, coiling and chamber arrangement, and position of apertures (ap). Bar scale = 200µm, unless indicated.

phology, multiple apertures, and flexible test. It occupies the muddy substrate at the base of upper marsh vegetation, in a habitat where environmental factors such as tidal changes and salinity fluctuations within a general hyposaline range may shape its development. The fragmentary wide distribution of the species may be associated with migratory waterbirds, which could facilitate its spread in mud attached to feathers and feet or in faeces. Further molecular and ecological research is essential to clarify its dispersal mechanisms and ecological role, particularly as an important indicator species in coastal ecosystems.

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Table 2. Presence and Absence of *Trochamminita irregularis* from estuaries around Australia.

Localities	Latitude and Longitude	States	Occurrence	References
Cocoa Creek	19.288° S, 147.005 °E	QLD	Absent	Horton et al., 2003; Woodroffe et al., 2005
Thomatis Creek, Barron River	16.846° S, 145.719° E	QLD	Absent	Haslett, 2001
South Alligator River	12.247° S, 132.403° E	NT	Absent	Wang & Chappell, 2001
Tuross Estuary and Coila Lake	35.736° S, 150.112° E	NSW	Absent	Strotz, 2003
Lake Illawarra	34.557° S, 150.841° E	NSW	Absent	Yassini & Jones, 1989
St George Basin	35.092° S, 150.596° E	NSW	Absent	Strotz, 2012
Smith Lake	32.251° S, 152.532° E	NSW	Absent	Strotz, 2015
Broken Bay estuary	33.566° S, 151.312° E	NSW	Absent	Albani, 1978
Minnamurra River	34.634° S, 150.847° E	NSW	Absent	Lal et al., 2020
Comerong Island	34.889° S, 150.735° E	NSW	Absent	Lal et al., 2020
Carama Inlet	34.993° S, 150.783° E	NSW	Absent	Lal et al., 2020
Currambene Creek	35.020° S, 150.667° E	NSW	Absent	Lal et al., 2020
Little Swanport Estuary	35.007° S, 150.215° E	TAS	Present	Callard et al., 2011
Tamar River and and Port Dalrymple	41.110° S, 146.798° E	TAS	Absent	Bell, 1996
Northern Spencer Gulf	33.007° S, 137.844° E	SA	Absent	Cann et al., 2002
Port Pirie	33.177° S, 138.008° E	SA	Absent	Cann et al., 2002
Lake Connewarre	38.226° S, 144.487° E	VIC	Absent	Bell, 1995
Mallacoota Inlet	37.563° S, 149.746° E	VIC	Absent	Bell & Drury, 1992
Barwon River estuary	38.292° S, 144.391° E	VIC	Absent	Parr, 1945
Gippsland LakeS, Victoria	38.091° S, 147.760° E	VIC	Present	Apthorpe, 1980
Mangrove Bay, Ningaloo Reef	21.962° S, 113.946° E	WA	Absent	Parker, 2009
Oyster Harbour	35.028° S, 117.884° E	WA	Absent	McKenzie, 1962
Hardy Inlet	33.541° S, 115.545° E	WA	Absent	Quilty, 1977
Murray River	34.300° S, 140.800° E	WA	Absent	Ostrognay & Haig, 2012
Leschenault Inlet and Collie River	33.283° S, 115.699° E	WA	Absent	Ostrognay & Haig, 2012; Revets, 2000; Tremblin et al., 2021
Wellstead Estuary	33.704° S, 118.396° E	WA	Absent	pers. obs.
Kalgan River	34.976° S, 117.741° E	WA	Absent	pers. obs.
Frankland River	33.957° S, 116.027° E	WA	Absent	pers. obs.
Walpole Inlet	34.969° S, 116.730° E	WA	Absent	pers. obs.
Mandurah marsh	32.548° S, 115.716° E	WA	Absent	pers. obs.
Hay River	34.970°S, 117.469°E	WA	Present	This study

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REFERENCES

Albani, A. D. (1978). Recent foraminifera of an estuarine environment in Broken Bay, New South Wales. *Marine and Freshwater Research, 29*, 355–398. <u>https://doi.org/10.1071/</u> MF9780355

Alve, E. (1999). Colonization of new habitats by benthic foraminifera: a review. *Earth-Science Reviews*, *46*, 167–185. https://doi.org/10.1016/S0012-8252(99)00016-1

Alve, E., & Goldstein, S. T. (2010). Dispersal, survival and delayed growth of benthic foraminiferal propagules. *Journal of Sea Research*, *63*, 36–51. <u>https://doi.org/10.1016/</u> j.seares.2009.09.003

Apthorpe, M. (1980). Foraminiferal distribution in the estuarine Gippsland Lakes System, Victoria. *Proceedings of the Royal Society of Victoria*, *91*, 207–232.

Avnaim-Katav, S., Garrett, E., Gehrels, W. R., Brown, L. N., Rockwell, T. K., Simms, A. R., Bentz, J. M., & MacDonald, G. M. (2023). Contemporary Salt-Marsh Foraminifera from Southern California and Implications for Reconstructing Late Holocene Sea- Level Changes. *Journal of Foraminiferal Research*, *53*, 157–176. https://doi.org/10.2113/gsjfr.53.2.157

Avnaim-Katav, S., Gehrels, W. R., Brown, L. N., Fard, E., & MacDonald, G. M. (2017). Distributions of salt-marsh foraminifera along the coast of SW California, USA: Implications for sea-level reconstructions. *Marine Micropaleontology*, *131*, 25–43. <u>https://doi.org/10.1016/</u> j.marmicro.2017.02.001

Bamford, M., Watkins, D., Bancroft, W., Tischler, G., & Wahl, J. (2008). *Migratory shorebirds of the East Asian-Australasian flyway: Population estimates and internationally important sites.* Wetlands International, Oceania.

Bell, K. N. (1995). Foraminiferans from Lake Connewarre, Victoria. *Victorian Naturalist, 112*, 228–234.

Bell, K. N. (1996). Foraminifera faunas of the River Tamar and Port Dalrymple, Tasmania: A preliminary survey. *Records of the Queen Victoria Museum Launceston*, *102*, 1–25.

Bell, K. N., & Drury, S. R. (1992). Foraminiferal fauna of Mallacoota Inlet, East Gippsland, Victoria. *Victorian Naturalist*, *109*, 7–16.

Birdlife Western Australia. (2024). *Birdwatching around Denmark*. <u>https://www.denmark.wa.gov.au/Profiles/</u> <u>denmark/Assets/ClientData/Denmark_Bird_Guide_2022.pdf</u>

Brann, B. C. (1969). Microfossils of the Trinity River Delta. In Houston Geological Society (Ed.), *Holocene geology of the Galveston Bay area* (pp. 118–126). Brearley, A. (2005). *Ernest Hodgkin's Swanland: Estuaries and Coastal Lagoons of Southwestern Australia*. University of Western Australia Press.

Bryant, S. (2002). *Conservation assessment of beach nesting and migratory shorebirds in Tasmania*. Nature Conservation Branch, Department Primary Industries Water and Environment.

Bunzel, K., Kattwinkel, M., & Liess, M. (2013). Effects of organic pollutants from wastewater treatment plants on aquatic invertebrate communities. *Water Research*, 47, 597–606. https://doi.org/10.1016/j.watres.2012.10.031

Bureau of Meteorology. (2024). *Average annual, seasonal and monthly rainfall maps*. Australian Government. <u>http://www.bom.gov.au/climate/maps/averages/rainfall/</u>

Callard, S. L., Gehrels, W. R., Morrison, B. V., & Grenfell, H. R. (2011). Suitability of saltmarsh foraminifera as proxy indicators of sea-level in Tasmania. *Marine Micropaleontology*, *79*, 121–131. <u>https://doi.org/10.1016/</u> J.MARMICRO.2011.03.001

Cann, J. H., Harvey, N., Barnett, E. J., Belperio, A. P., & Bourman, R. P. (2002). Foraminiferal biofacies eco-succession and Holocene sea-levels, Port Pirie, South Australia. *Marine Micropaleontology*, 44, 31–55. <u>https://doi.org/10.1016/</u> <u>S0377-8398(01)00036-6</u>

Clague, J. J., Bobrowsky, P. T., & Hamilton, T. S. (1994). A sand sheet deposited by the 1964 Alaska tsunami at Port Alberni, British Columbia. *Estuarine, Coastal and Shelf Science, 38*, 413–421. https://doi.org/10.1006/ecss.1994.1028

Coughlan, N. E., Kelly, T. C., Davenport, J., & Jansen, M. A. (2017). Up, up and away: Bird-mediated ectozoochorous dispersal between aquatic environments. *Freshwater Biology*, *62*(4), 631–648. <u>https://doi.org/10.1111/fwb.12894</u>

Culver, S. J., & Horton, B. P. (2005). Infaunal marsh foraminifera from the Outer Banks, North Carolina, U.S.A. *Journal of Foraminiferal Research*, *35*, 148–170. <u>https://</u> <u>doi.org/10.2113/35.2.148</u>

Culver, S. J., Leorri, E., Mallinson, D. J., Corbett, D. R., & Shazili, N. A. M. (2015). Recent coastal evolution and sea-level rise, Setiu Wetland, Peninsular Malaysia. *Palaeogeography, Palaeoclimatology, Palaeoecology, 417*, 406–421. <u>https://</u> doi.org/10.1016/j.palaeo.2014.10.001

Cushman, J. A., & Brönnimann, P. (1948). Some new genera and species of foraminifera from brackish water of Trinidad. *Contribution to Laboratory for Foraminiferal Research*, 24, 15–21. Debenay, J.-P., Guiral, D., & Parra, M. (2002). Ecological factors acting on the microfauna in mangrove swamps: The case of foraminiferal assemblages in French Guiana. *Estuarine, Coastal and Shelf Science, 55*, 509–533. <u>https://doi.org/10.1006/ecss.2001.0906</u>

Debenay, J.-P., Marchand, C., Molnar, N., Aschenbroich, A., & Meziane, T. (2015). Foraminiferal assemblages as bioindicators to assess potential pollution in mangroves used as a natural biofilter for shrimp farm effluents (New Caledonia). *Marine Pollution Bulletin, 93*, 103–120. <u>https://doi.org/</u> 10.1016/j.marpolbul.2015.02.009

Dreher, C. (2006). *Modern foraminiferal biofacies within a transgressive saline influenced deltaic headland, South-Central Louisiana*. University of New Orleans Theses and Dissertations.

Fatela, F., Moreno, J., Leorri, E., & Corbett, R. (2014). High marsh foraminiferal assemblages' response to intra-decadal and multi-decadal precipitation variability, between 1934 and 2010 (Minho, NW Portugal). *Journal of Sea Research*, *93*, 118–132. https://doi.org/10.1016/j.seares.2013.07.021

Finger, K. L. (2018). Tsunami-generated rafting of foraminifera across the North Pacific Ocean. *Aquatic Invasions*, *13*, 17–30. <u>https://doi.org/10.3391/ai.2018.13.1.03</u>

Fürstenberg, S., Gründler, N., Meschner, S., & Frenzel, P. (2017). Microfossils in surface sediments of brackish waters on the west coast of South Africa and their palaeoecological implications. *African Journal of Aquatic Science*, *42*, 329–339. https://doi.org/10.2989/16085914.2017.1406326

Garrett, E., Gehrels, W. R., Hayward, B. W., Newnham, R., Gehrels, M. J., Morey, C. J., & Dangendorf, S. (2022). Drivers of 20th-century sea-level change in southern New Zealand determined from proxy and instrumental records. *Journal of Quaternary Science*, *37*, 1025–1043. <u>https://doi.org/10.1002/</u> jqs.3418

Gehrels, W. R., Callard, S. L., Moss, P. T., Marshall, W. A., Blaauw, M., Hunter, J., Milton, J. A., & Garnett, M. H. (2012). Nineteenth and twentieth century sea-level changes in Tasmania and New Zealand. *Earth and Planetary Science Letters*, *315–316*, 94–102. <u>https://doi.org/10.1016/</u> j.epsl.2011.08.046

Guy-Haim, T., Hyams-Kaphzan, O., Yeruham, E., Almogi-Labin, A., & Carlton, J. T. (2017). A novel marine bioinvasion vector: Ichthyochory, live passage through fish. *Limnology and Oceanography Letters*, 2, 81–90. <u>https://doi.org/10.1002/</u> <u>lol2.10039</u>

Haller, C., Smith, C. G., Hallock, P., Hine, A. C., Osterman, L. E., & McCloskey, T. (2019). Distribution of modern saltmarsh foraminifera from the Eastern Mississippi Sound, U.S.A. *Journal of Foraminiferal Research*, 49, 29–47. https://doi.org/ 10.2113/gsjfr.49.1.29 Hansen, B. D., Healey, C., Sullivan, D., & Weller, D. R. (2024). Waterbird and migratory shorebird monitoring in the Gippsland Lakes. *Proceedings of the Royal Society of Victoria*, *136*, RS23030. https://doi.org/10.1071/RS23030

Haslett, S. K. (2001). The palaeoenvironmental implications of the distribution of intertidal foraminifera in a tropical Australian estuary: a reconnaissance study. *Australian Geographical Studies*, *39*, 67–74. <u>https://doi.org/10.1111/</u> <u>1467-8470.00130</u>

Hawkes, A. D., Horton, B. P., Nelson, A. R., & Hill, D. F. (2010). The application of intertidal foraminifera to reconstruct coastal subsidence during the giant Cascadia earthquake of AD 1700 in Oregon, USA. *Quaternary International*, *221*, 116–140. https://doi.org/10.1016/ j.quaint.2009.09.019

Hayward, B. W., Hollis, C. J., & Grenell, H. (1994). Foraminiferal associations in Port Pegasus. Stewart Island, New Zealand. *New Zealand Journal of Marine and Freshwater Research*, 28, 69–95. <u>https://doi.org/10.1080/</u> 00288330.1994.9516597

Horton, B. P., Larcombe, P., Woodroffe, S. A., Whittaker, J. E., Wright, M. R., & Wynn, C. (2003). Contemporary foraminiferal distributions of a mangrove environment, Great Barrier Reef coastline, Australia: implications for sea-level reconstructions. *Marine Geology*, *198*, 225–243. <u>https:// doi.org/10.1016/S0025-3227(03)00117-8</u>

Jennings, A. E., Nelson, A. R., Scott, D. B., & Aravena, J. C. (1995). Marsh foraminiferal assemblages in the Valdivia estuary, south-central Chile, relative to vascular plants and sea level. *Journal of Coastal Research*, *11*, 107–123.

King, D. J. (2021). A Re-evaluation of the foraminiferal genus Trochamminita (Cushman and Brönnimann, 1948) in New Zealand and a description of Pseudotrochamminita malcolmi (new genus, new species). *Journal of Foraminiferal Research*, *51*, 308–317.

Lal, K. K., Bonetti, C., Woodroffe, C. D., & Rogers, K. (2020). Contemporary distribution of benthic foraminiferal assemblages in coastal wetlands of south-eastern Australia. *Estuarine, Coastal and Shelf Science, 245*, Article 106949. https://doi.org/10.1016/j.ecss.2020.106949

Laut, L., Clemente, I., Martins, M. V. A., Frontalini, F., Raposo, D., Belart, P., Habib, R., Fortes, R., & Lorini, M. L. (2017). Benthic Foraminifera and Thecamoebians of Godineau River Estuary, Gulf of Paria, Trinidad Island. *Anuário do Instituto de Geociências*, 40, 118–143. <u>https://doi.org/</u> 10.11137/2017_2_118_143

Laut, L. L. M., Martins, M. V. A., Frontalini, F., Belart, P., Santos, V. F., Lorini, M. L., Fortes, R. R., Silva, F. S., Vieira, S. S. S., & Souza-Filho, P. W. M. (2016). Biotic (foraminifera and thecamoebians) and parameters as proxies for indication of the environmental heterogeneity in Caeté River Estuary, Amazon Coast, Brazil. *Journal of Sedimentary Environments*, 1, 1–16. https://doi.org/10.12957/jse.2016.21264 Lehmann, G. (2000). Vorkommen, Populationsentwicklung, Ursache fleckenhafter Besiedlung und Fortpflanzungsbiologie von Foraminiferen in Salzwiesen und Flachwasser der Nordund Ostseeküste Schleswig-Holsteins [Doctoral dissertation]. University of Kiel, Germany.

Lübbers, J., & Schönfeld, J. (2018). Recent saltmarsh foraminiferal assemblages from Iceland. *Estuarine, Coastal and Shelf Science, 200*, 380–394. <u>https://doi.org/10.1016/</u> j.ecss.2017.11.019

Lukatelich, R. J., Schofield, N. J., & McComb, A. J. (1984). *The nutrient status of Wilson Inlet 1982-1983*. Department of Conservation and Environment.

Lukatelich, R. J., Schofield, N. J., & McComb, A. J. (1987). Nutrient loading and macrophyte growth in Wilson Inlet, a bar-built southwestern Australian estuary. *Estuarine, Coastal and Shelf Science, 24*, 141–165. <u>https://doi.org/10.1016/</u> 0272-7714(87)90062-x

Martins, M. A., Hohenegger, J., Frontalini, F., Sequeira, C., Miranda, P., Rodrigues, M. D. C., ... Rocha, F. (2019). Foraminifera check list and the main species distribution in the Aveiro lagoon and adjacent continental shelf (Portugal). *Journal of Sedimentary Environments*, *4*, 1–52. <u>https://</u> <u>doi.org/10.12957/jse.2019.39308</u>

McKenzie, K. G. (1962). A record of Foraminifera from Oyster Harbour, near Albany, Western Australia. *Journal of the Royal Society of Western Australia*, 45, 117–133.

Milker, Y., Horton, B. P., Nelson, A. R., Engelhart, S. E., & Witter, R. C. (2015). Variability of intertidal foraminiferal assemblages in a salt marsh, Oregon, USA. *Marine Micropaleontology*, *118*, 1–16. <u>https://doi.org/10.1016/</u> j.marmicro.2015.04.004

Milker, Y., Schönfeld, J., Meng, S., Wrozyna, C., Schneppmüller, M., & Schmiedl, G. (2023). Are They Everywhere? –Benthic Foraminifera From Saline Springs in Central Germany. *Journal of Foraminiferal Research*, *53*, 286–304. https://doi.org/10.2113/gsjfr.53.4.286

Moreno, D. P. (2004). Distribuição dos foraminíferos recentes associados à vegetação na faixa estuarina do Rio Itapanhaú, Bertioga, São Paulo. São Paulo State University.

Murray, J. W. (2014). *Ecology and palaeoecology of benthic foraminifera*. Routledge. <u>https://doi.org/10.4324/</u> <u>9781315846101</u>

Ostrognay, D. B., & Haig, D. W. (2012). Foraminifera from microtidal rivers with large seasonal salinity variation, southwest Western Australia. *Journal of the Royal Society of Western Australia*, *95*, 137–153. Ozarko, D. L., Patterson, R. T., & Williams, H. F. L. (1997). Marsh foraminifera from Nanaimo, British Columbia (Canada); implications of infaunal habitat and taphonomic biasing. *The Journal of Foraminiferal Research*, *27*, 51–68. https://doi.org/10.2113/gsjfr.27.1.51

Parr, W. J. (1945). Recent Foraminifera from Barwon Heads, Victoria. *Proceedings of the Royal Society of Victoria*, 56, 189–218.

Pennings, S. C., & Bertness, M. D. (2001). Salt marsh communities. *Marine Community Ecology*, *11*, 289–316.

Quilty, P. G. (1977). Foraminifera of Hardy Inlet, southwestern Australia. *Journal of the Royal Society of Western Australia*, *59*, 79–90.

Ranasinghe, R., & Pattiaratchi, C. (1999). The seasonal closure of tidal inlets: Wilson Inlet—a case study. *Coastal Engineering*, *37*, 37–56. <u>https://doi.org/10.1016/s0378-3839(99)00007-1</u>

Reed, D. J., Davidson-Arnott, R., & Perillo, G. M. (2009). Estuaries, coastal marshes, tidal flats and coastal dunes. *Geomorphology and Global Environmental Change*, *30*, 130–157. https://doi.org/10.1017/CBO9780511627057.006

Resig, J. M. (1974). Recent foraminifera from a landlocked Hawaiian lake. *The Journal of Foraminiferal Research*, *4*, 69–76. <u>https://doi.org/10.2113/gsjfr.4.2.69</u>

Revets, S. A. (2000). Foraminifera of Leschenault Inlet. *Journal of the Royal Society of Western Australia*, *83*, 365–375.

Riedel, F., Kossler, K., Tarasov, P., & Wünnemann, B. (2011). A study on Holocene foraminifera from the Aral Sea and West Siberian lakes and its implication for migration pathways. *Quaternary International*, 229, 105–111. <u>https://doi.org/</u> 10.1016/j.quaint.2010.03.009

Robinson, M. M., & McBride, R. A. (2006). Benthic foraminifera from a relict flood tidal delta along the Virginia/ North Carolina Outer Banks. *Micropaleontology*, *52*, 67–80. https://doi.org/10.2113/gsmicropal.52.1.67

Ross, P., Minchinton, T., & Ponder, W. (2009). *The ecology of molluscs in Australian saltmarshes*. CSIRO Publishing.

Saunders, J. B. (1957). Trochamminidae and certain Lituolidae (Foraminifera) from the recent brackish-water sediments of Trinidad, British West Indies. *Smithsonian Miscellaneous Collections*, 134, 1–16.

Semensatto, D. (2020). A key to the identification of agglutinant and monothalamous foraminifera from Brazilian mangroves. *Ocean and Coastal Research, 68*, e20297. <u>https://doi.org/10.1590/s2675-28242020068297</u>

Strotz, L. C. (2003). Holocene Foraminifera from Tuross Estuary and Coila Lake, South Coast, New South Wales: A preliminary study. *Proceedings of the Linnean Society of New South Wales, 124*, 163–182.

Strotz, L. C. (2012). Foraminiferal fauna and biotopes of a barrier estuary system, St Georges Basin, New South Wales, Australia. *Journal of Foraminiferal Research*, *42*, 369–382. https://doi.org/10.2113/GSJFR.42.4.369

Strotz, L. C. (2015). Spatial patterns and diversity of foraminifera from an intermittently closed and open lagoon, Smith Lake, Australia. *Estuarine, Coastal and Shelf Science, 164*, 340–352. <u>https://doi.org/10.1016/J.ECSS.2015.07.048</u>

Tremblin, C. M., Holzmann, M., Parker, J. H., Sadekov, A., & Haig, D. W. (2021). Invasive Japanese foraminifera in a southwest Australian estuary. *Marine and Freshwater Research*, *73*, 328–342. https://doi.org/10.1071/MF21254

Twomey, L., & Thompson, P. (2001). Nutrient limitation of phytoplankton in a seasonally open bar-built estuary: Wilson Inlet, Western Australia. *Journal of Phycology*, *37*, 16–29. https://doi.org/10.1046/j.1529-8817.1999.014012016.x

Verlaak, Z. R., & Collins, L. S. (2021). Environmental controls on the distribution of modern benthic foraminifera in the Florida Everglades and their use as paleoenvironmental indicators. *Journal of Foraminiferal Research*, *51*, 182–209. https://doi.org/10.2113/gsjfr.51.3.182 Verlaak, Z. R. F. (2019). Benthic Foraminiferal Assemblages from Marshes and Mangroves in the Everglades (South Florida, USA) and Their Application as Proxies for Habitat Shifts Due to Sea Level Rise [Doctoral dissertation]. Florida International University.

Vernberg, F. J. (1993). Salt-marsh processes: a review. Environmental Toxicology and Chemistry: An International Journal, 12, 2167–2195.

Wang, P., & Chappell, J. (2001). Foraminifera as Holocene environmental indicators in the South Alligator River, northern Australia. *Quaternary International*, *83*, 47–62. https://doi.org/10.1016/S1040-6182(01)00030-1

Williams, S., Garrett, E., Moss, P., Bartlett, R., & Gehrels, W. R. (2021). Development of a training set of contemporary saltmarsh Foraminifera for Late Holocene sea-level reconstructions in southeastern Australia. *Open Quaternary*, *7*, 1–29. <u>https://</u><u>doi.org/10.5334/oq.93</u>

Woodroffe, S. H., Horton, B. P., Larcombe, P., & Whittaker, J. E. (2005). Intertidal mangrove foraminifera from the central Great Barrier Reef shelf, Australia: Implications for sea-level reconstruction. *Journal of Foraminiferal Research*, *35*, 259–270. https://doi.org/10.2113/35.3.259

Yassini, I., & Jones, B. G. (1989). Estuarine foraminiferal communities in Lake Illawarra. *Proceedings of the Linnean Society of New South Wales, 110, 229–266.*